Synchronization of Estrus in Naeimi Ewes Following Treatment with Progrestagens or Prostaglandin $F_{2\alpha}$

A. M. Homeida, A. I. AL-Mubarak And Y. M. AL-Yousef^{*}

Camel Research Centre *College of Agricultural and Food Sciences King Faisal University, AL-Ahsa, Saudi Arabia

Abstract :

The efficacy of using progesterone releasing interavaginal device (PRID), PRID without estradiol benzoate, veramix sheep sponge and prostaglandin $F_{2\alpha}$ in inducing and synchronizing estrus were evaluated in Naeimi ewes. All four treatments were successfully able to synchronize estrus. The interval to induce estrus was significantly (P<0.05) shorter and estrous intensity was significantly higher in PRID-treated group compared to other treatment groups. Serum progesterone values during subsequent estrous cycle and pregnancy rate were similar in all treatment groups.

The results indicated that progestagens and prostaglandin F2 α can effectively be used in management of reproduction in Naeimi sheep.

Introduction :

Inspite of the numerical and economical importance of the Awassi sheep in Saudi Arabia, few planned research investigations were conducted to study the various factors which affecting their reproduction.

The study of reproduction in Awassi breed received considerable attentions in neighbouring countries (Hossamo *et al.*, 1985 in Syria; Eliya *et al.*, 1972 in Iraq; Finci 1975 in Palestine; Khalil *et al.*, 1972 in Lebanon). Similar information on the indigenous Naeimi sheep which is a branch of the Awassi breed of Saudi Arabia is scant.

Estrus synchronization has become a vital instrument in the management of reproduction in domestic farm animals (Motlomelo *et al.*, 2002; Fonesca *et al.*, 2008). Its benefits include a planned breeding programme and a reduction in labor costs in terms of estrus detection and care of the newborn. Estrus synchronization has also become an important tool for embryo transfer.

The objective of this study was to evaluate the efficacy of progestagens and prostaglandin F2 α in synchronizing estrus in Awassi (Naeimi) sheep, and to record pregnancy thereafter.

Materials and Methods : Animals and treatments :

Eighty mature (aged 2-4 years and weighing 30-40kg) (Naeimi sheep) were used in the study. They were housed in pens under conditions of natural day length and temperature. They were fed on a roughage mixture composed of 50% green fodder (*Medicago sativa*) and 50% Alfalfa hay offered *ad libititum*. Water and salt licks was available at all times. All ewes were checked for estrus when they stand for mounting by a ram this is termed day 0 of the estrous cycle. Animals were allocated randomly and equally to groups of 20 animals each:

- Group A: ewes were injected with PGF2α (Dinoprost Tromethamine, Upjohn Company, Kalamazoo, USA; 10mg i.m) at day 0 and then at 11-day intervals.
- Group B: ewes were treated with progesterone releasing intravaginal device (PRID). The PRID used had a progesterone content of 1.55g, and a capsule containing 10mg of estradiol benzoate attached the coil.
- Group C : ewes were treated with PRID after removal of the capsule of estradiol benzoate from to the coil.
- Group D: ewes were treated with Veramix sheep sponges (Upjohn Ltd. Fleming Way, Crawley, Sussex, England) containing 60mg 6methyl-17-acetoxy-progesterone. Sponges and PRIDS were inserted in the anterior vagina at day 0, and were removed 14 days later.

Ewes were exposed three times daily (0600, 1200 and 1800h) post treatment to aproned rams. The duration of standing estrus was notified by immobility when mounted by a male, while the length of the estrous cycle was regarded as the interval from the end of estrus to the beginning of the next one in the same ewe. The intensity of estrus was scored on a scale from zero to 3 as the degree of expression of restlessness, standing to be mounted, vocalization, and swelling of vulva and mucus discharge. Animals were then bred by natural service and pregnancy was recorded.

Collection of Blood Samples :

After completion of one normal cycle, blood sampling was commenced on day 2 of estrous cycle. Jugular blood (5ml) is obtained once daily by direct venepuncture using 23 gauge needles. From day 12 through to day of estrus, samples were taken at two hour intervals between 07:00 and 17:00 hours.

All blood samples were collected into chilled, heparinised tubes, stored in ice for a short period and then centrifuged at 1500g for 10 minutes. Plasma was separated and stored at -30° C until analysis.

Radioimmunoassay of hormones :

Plasma progesterone was measured by the radioimmunoassay (RIA) method described by Homeida (1986), and Homeida & Al-Eknah (1992). Progesterone antibody was raised in raised in rabbits against Progesterone-11-succiny-bovine serum albumin, and used at a final dilution of 1:7000; cross-reactions were 100% with progesterone and <0.1% with corticosterone, desoxycorticosterne and ketocrticosterone. The intra- and inter-assay coefficients of variation were 5.3% and 9.1%, respectively.

Statistical analysis :

Comparisons of hormone concentrations over time were made by ANOVA, taking of repeated measures from same animal (Steele and Torrie 1988).

Results :

Estrual responses following treatment with PRIDs, veramix sponge or $PGF_{2\alpha}$ treatment in Naeimi ewes are presented in table (1) Neither PRIDs nor sponge were lost during the treatment periods, Estrous was synchronized in most of ewes by the four treatment methods. The interval to onset estrus were significantly (P<0.05) shorter in group B (44.2 hour) than in group C (52.6 hour) or group D (77.6 hour) following removal of the devices. The interval in group B was similar to group A (41.3 hour) following the second PGF_{2a} injection. The duration of subsequent estrous cycle was 16.3, 16.5, 16.2 and 17.3 days in groups A, B, C and D, respectively.

The estrus intensity score was significantly (P<0.05) higher in group B compared to other groups.

Number of animals in estrous and number of pregnant animals were higher though not significant in group B compared to other groups.

Mean serum progesterone concentrations are presented in table (2). Values of serum progesterone concentrations were significantly (P<0.05) higher in group B compared to other groups during device insertion. Serum progesterone concentrations increased steadily following first PGF2 α treatment or device insertion. The pattern of the hormone was similar during the subsequent estrous cycle but tended to be higher in group B, C and D compared to group A.

Synchronization of Estrus in Naeimi Ewes Following
--

Tabel (1) : Estrual responses following treatment with PRIDs Veramix sponge or PGF $_{2u}$ treatments in Naeimi ewes	ing treatment with PF	SIDs Veramix spong	je or \mathbf{PGF}_{2a} treatment	s in Naeimi ewes
Parameter	Group A (PGF _{2a} treated)	Group B (PRID Treated)	Group C (PRID with- estradiol treatment)	Group D (Veramix reated)
Number of treated animals	20	20	20	20
Number of animals lost device	-	0	0	0
Number in estrus	16	18	14	15
Mean interval to estrus (hour)	41.3±2-1	44.2± 1.2	52.6± 1.4*	77.6± 3.1*
Mean estrus intensity (score 12)	6 ± 0.1	$12 \pm 0.3*$	6 ± 0.2	6 ± 0.1
Duration of estrous cycle (days)	16.2 ± 0.3	16.5±0.3	16.2±0.2	17.3±0.3
Number of pregnant animals	13	16	12	12
* P<0.05, significantly different from their counterparts.	counterparts.			

2

treatment or vaginal device insertion and removal in Naeimi ewes.					
Days following device insertion or first PGF _{2a} injection	Group A	Group B	Group C	Group D	
2	0.5 ± 0.2	2.1 ±0.2	1.5 ± 0.2	1.2 ± 0.2	
4	1.1 ± 0.2	4.4± 0.3*	2.5 ± 0.2	2.1 ± 0.2	
6	2.4 ± 0.3	6.1±0.4*	3.1 ± 0.2	3.3 ± 0.3	
8	3.9 ± 0.3	6.6 ± 0.4	3.9 ± 0.3	4.2 ± 0.3	
10	4.2 ± 0.3	7.4±0.4*	4.1 ± 0.3	5.1 ± 0.3	
12	2.1 ± 0.2	7.3±0.4*	4.1 ± 0.3	5.3 ± 0.4	
14		7.1±0.3*	4.6 ± 0.3	5.5 ± 0.4	
Days following device removal or second $PGF_{2\alpha}$ injection 2	0.4 ± 0.1	0.5 ± 0.1	0.4 ± 0.1	0.3 ± 0.1	
4	0.4 ± 0.1	1.1 ± 0.1	1.4± 0.2	1.2 ± 0.1	
6	.25 ± 0.2	2.4 ± 0.2	2.6 ± 0.2	2.5 ± 0.2	
8	1.1 ± 0.3	4.5 ± 0.3	5.6 ± 0.3	5.4 ± 0.3	
10	4.3 ± 0.3	5.2 ± 0.3	5.3 ± 0.3	5.1 ± 0.4	
12	3.6 ± 0.3	3.2 ± 0.3	3.1 ± 0.3	2.9 ± 0.3	
14	2.3 ± 0.2	2.1 ± 0.2	2.4 ± 0.2	2.3 ± 0.2	
16	0.6 ± 0.1	0.5 ± 0.1	6.4 ± 0.1	0.3 ± 0.1	
18	0.2± 0.1	0.4 ± 0.1	0.3 ± 0.1	0.2 ± 0.1	

Table (2)
Mean (\pm SD) serum progesterone concentration (ng/ml) during PGF _{2a}
treatment or vaginal device insertion and removal in Naeimi ewes.

* P< 0.05

Discussion :

The four estrus synchronizing methods have effectively synchronized estrus in Naeimi sheep. Comparable efficacy achieved by progestagens in synchronizing estrus has been reported in Syrian Awassi ewes (Zarkawi *et al.* 1999) and other breeds of sheep (kusakari *et al.*, 1991; Das et al. 2001; Mutiga and Mukasa – Mugerwa 1992) and goats (Fonseca *et al.* 2008). The mean interval to estrous was shorter and the intensity of estrus was higher in animals treated with PRID than in other animal groups. This is probably due to estrogen being incorporated in the PRID. Enhanced estrus expression and behavior was noticed when estradiol was given in conjunction with PRID in cows (Walton and king 1984, Narasimha & suryapakasam 1991)

Furthermore, (coetzer *et al.*, 1988) showed that removal of estradiol benzoate from a PRID device resulted in poor estrus response.

The degree of synchrony and interval to estrus following the second PGF2 α injection were consistent with earlier reports in different breeds of sheep (Acritopoulou *et al.*, 1978; Haesign 1978; Oydiji *et al.*, 1990).

Although fertility trials were not assessed critically in this experiment but pregnancy rate was similar with all synchronized methods. Pregnancy rate was high in sheep following progestagen (Das *et al.*, 2004) and prostagland via F2 α (Gardenas et al. 1993) treatments.

Administration of PRID resulted in significantly higher progesterone concentration compared to PRID without estradiol benzoate or veramix sponge. This is probably due to modulating effect of estradiol on the absorption of progesterone from vagina in the PRID group and the difference in physicochemical properties between the natural progesterone contained in PRID and the synthetic progesterone contained in veramix sponge.

The pattern of progesterone following the device removal or following the second PGF2 α , injection were similar to the natural estrous cycle in sheep and cattle (Cardenas *et al.*, 1993; Broadbent *et al.*, 1993).

In conclusion, the present study indicated that intravaginal progestagen device or prostaglandin F2 α can be used effectively for induction and synchronization of estrus in Naeimi sheep. Behavioral estrus was better expressed in the PRID group. Pregnancy rate was similar in all treatment methods.

Acknowledgements:

The authors thank the Deanship of scientific research, KFU for financial support and Dr. Mohammed Al-Bachiet, for technical assistance.

References :

- 1. Acritopoulou, S., Hare sign, W. and Lamming, G.E. Time of ovulation in ewes after treatment with a $PGF_{2\alpha}$ analogue. J. Report. Fertil. 54:189-191 (1978).
- 2. Broadbent P.J. Stewart M. and Dolman D. F. (1991) Recipient management and embryo transfer. Theriogenology 35, 125-139.
- 3. Coetzer W. A. Van Nie Kerk C. H and Esterhuyse A. J. (1988) the use of progesterone releasing intravaginal device (PRID) as a synchronizing agent in dairy heifers. Proc. 11th Int. Cong. Anim. Report. A. I. 443.
- Das G. K. Naqi S.M.K Gulyani R., Pareek S. R Narula H. K. and Mittal J. P. (2004) Estrus induction and fertility response in a cycling Awassi – Malpura ewes treated with progesterone and PMSG in tropical climate. Indian. J. of Anim. Sci 74, 713-717.
- 5. Das GK, Naqvi S M K, Gulyani R pareek ST and Mitten J P. 2001 Estrus response and fertility in postpartum ewes using progesterone and PMSG in tropical environment. Indian Journal of Animal Sciences 71:1124-26.
- Eliya, J. and Juma, K. H and Al- Shabibi, M. A. (1972). A note on the composition and properties of Awassi sheep milk. Egypt j. Anim. Prod. 12 (1): 15-45.
- 7. Finci, M. (1975). The improvement of Amass breast sheep in Palestine, Bulletin, Research Council, Palestine, B, 6, 1.
- Fonseca J.F. Torres C.A.A., Santis A.D.F., Maffili V.V. Amorim L.S., and moraes E.A. (2008). Progesterone and behavioral features when estrous is induced in Alpine goats. Anim. Reprod. Sci. 103. 366-373.
- Gardenas H., Mc Clure K. E. and Pope W.F (1993) Luteal function and blastoajst development in ewes following treatment with PG F₂x and GnRH. Theriogenology 40, 865-872.
- Haresign, W. Ovulation control in the sheep. In: Cryghton, D.B., Haynnes, N.B., Fox croft, G.R. and Lamming, G.E. (eds), Control of Ovulation, Butterworth, London, 1978, pp.435-451.
- Homeida A. M. (1986) use of Spiro no lactones to investigate thd role of testosterone secretion during lutreolysis in the goat. J. Reprod. Fret. 76-153-157.
- 12. Homeida A.M. and Al-Eknah M.M. (1992) inhibition of brutal function by oxytocin antagonist in goats (Capra circus). J. Reprod. Fret. 94,249-285.
- Hossamo, H. E., J. B. Owen, and Farid, F. A. (1985). The genetic improvement of syriam Awash sheep with special reference to milk production J. Agric. Sci. Camb. 105, 327-337.

- Khalil, K., Choueiri, E., Fox., O. W., Badawi, S. Harrison, W. H., and Mata, A. (1972). Milk production Fro a selection flock of Awassi sheep. 4th Science, Absrack, 52.
- 15. Kusakari N, kisi Kand O'Hara M. 1991. Effect of intravaginal sponges impregnated with varying doses of progesterone on estrus response in seasonally anoestrus ewes. Japanese journal of Animal Reproduction 37:27-31.
- 16. Montlomelo K.C. Greyling J.P.C. Schwalbach L.M.J. (2002). Synchronization of estrus in goats: the use of different progestagen treatments. Small Rumin. Res. 45. 45-49.
- 17. Mutiga E R and Mukasa –Mugerwa E. 1992. Effect of the method of estrus synchronization and PMSG dosage on estrus twinning in Ethiopian Menze Sheep. Theriogenology 38:727-34.
- 18. Narasimha Rao A. V. and suryaprakasam T. B. (1991) Induction of synchronized estrus and fertility in anestrous zebu x Taurus crossbred cows. Therogenolgy 36, 123-128.
- 19. Oyediji G. O. Akusu M. O. and Egbunke G. N. (1990) comparative studies of effectiveness of Sil- estrus implants, Veramix sheep sponges and prostaglandin $F_{2\alpha}$ in synchronizing estrus in West African dwarf sheep. Therogenology 34, 613-618.
- 20. Steele R.G. and Torrie J.H.(1980). Principles and procedures of statistics. A Biometrical Approach. McGraw-Hill Book Co. New York 131-197.
- Walton J. S and King G. T(1984) the effect of progesterone treatment on estradiol induced estrous behavior in ovariectomized cows Proc. 10th int. Cong. Anim. Reprod. and A. I. Champaign pp229-230.
- 22. Zarkawi M, Al-Merestani M R and Wardeh M F. 1999. Induction of synchronized oestruz and early pregnancy diagnosis in Syrian Awassi ewes, outside the breeding season. Small Ruminant Research 33:99-102.



إحداث تزامن الشبق في النعاج النعيمي بواسطة البروجستاجين والبروستجلاندين F_{2a}

عبدالقادر حميده وعادل المبارك و يوسف اليوسف*

مركز أبحاث الجمال * كلية العلوم الزراعية والأغذية جامعة الملك فيصل ، الأحساء ، المملكة العربية السعودية

الملخص:

لقد تم تقيم مقدرة أحداث تزامن الشبق في النعاج النعيمي بواسطة الحشوة البروجستينة أو الحشوة البروجستينية بدون الاستروجين أو الحشوة الأسفنجية الفيرموكس أو البروجستجلاندين – ف – الفا . لقد تم أحداث تزامن الشبق بنجاح بواسطة الأدوية المستخدمة . لقد تم أحداث الشبق في وقت اقصر كما وان ظواهر الشبق كان بصورة أوضح في النعاج المعالجة بالحشوة البروجستينة بالمقارنة مع بقية المعالجات ولقد كان تركيز البروجستيرون في الدورة اللاحقة للمعالجة والحمل متشابه في المعالاجات الأربعات . هذه النتائج تشير إلى مقدرة البروجستاجين والبروجستلاندين في إدارة النواحي التناسلية في نعاج النعيمي.